

Vaccine composition containing transforming growth factor alpha (TGF α). It use in malignant diseases therapy.

Technical field:

- 5 This invention relates to human medicine, and especially to therapeutic vaccines comprising TGF α ; and particularly provides vaccine compositions useful in cancer immunotherapy.

Prior art

- 10 TGF α is a 50 amino acids polypeptide; it was isolated originally from the conditioned medium by retrovirus-transformed cells. By reason of binding the same receptor, TGF α is considered a member of the EGF family molecules, which comprise structural and functional related proteins. But antibodies anti-EGF do not recognize TGF α (Todaro et al. (1976), Nature 264, 26-31, which leads to the issue that they are two immunological different entities.

- 15 Other members of EGF family are amphiregulin (AR), criptol (CR1), heparin-binding EGF, betacellulin, epiregulin. Poxvirus family also includes EGF related proteins, the best characterized is the vaccinia virus growth factor (VGF).

- By the reason of binding and triggering EGF-R, all the before mentioned molecules are considered as ligands of this receptor and they play a role in the regulation of normal and neoplastic cells growth.

- 20 EGF-R is a glycoprotein of 170 kD with kinase activity in tyrosine residues whose gene has been cloned and sequenced. These proteins have shown structural homology with the product of v-erb-B oncogen, what is an evidence of the relationship between the neoplastic transformation process and this molecules (Heldin C.H. (1984), Cell 37, 9-20.)

- 25 TGF α is synthesized as a transmembrane precursor (pro-TGF α) with 160 amino acids. Mature TGF α , a soluble polypeptide with 50 amino acids, is released by proteolytic cleavage. Human TGF α (hTGF α) shows 43% amino acid sequence identity with human EGF (hEGF) and 93% with mouse or rat TGF α . Besides, their biological effects are not species-specific.

- it has shown the ability of TGF α to regulate the proliferation, migration and differentiation of cultured cells (Carpenter and Wahl. (1990), Springer-Verlag, Berlin, pp.69-171.)

- 30 TGF α is the most widespread ligand of EGF-R. It is expressed in normal tissues during the embryogenesis and in normal and tumor tissues in adults. However, not major pathological defects are observed in TGF α knockout mice and these mice are viable and fertile (Bruce Mann and cols. (1993), Cell, 73, 249-261.).

- 35 In the tumorigenesis process, the deregulation of paracrine and autocrine processes of EGF-R activation is due to the up-regulation of growth factor expression or the high synthesis or mutation of their receptor.

- The over-expression of these receptors in some kinds of neoplasia, mainly of epithelial origin, has been shown to be valuable as indicators of a bad prognosis in cancer.

- 40 The induction of TGF α expression is a frequent event during malignant transformation. over-expression of this molecule in neoplasia of epithelial origin has been reported, this is the case of breast, lung, brain, liver, prostate, bladder, gastrointestinal tract, colon, ovaryia, vulvae and endocrine tissues, .

- 45 Although the mechanism by which TGF α induces tumorigenicity remains unknown, there are some reports that correlate the growth factor over-expression with tumor degree, survival of patient and other tumoral markers. Besides, some researchers have demonstrated their relationship with other oncogenes like c-myc in hepatocarcinomas. TGF α also constitutes a target of the von Hippel-Lindau tumor suppressor gene (VHL) (Sumarized in Lee et al. (1996), Growth Factors and Cytokines in Health and Disease, Volume 1B, 277-318.).

- 50 Although TGF α and EGF bind the same receptor with comparable affinities, TGF α is generally more potent than EGF, and in some contexts, its effects have been described as stronger and/or more prolonged (Barrandon and Green (1987), Cell 50, 1131-1137). It has been reported that in the case of an internalized TGF α /EGF-R complex, TGF α and EGF-R were preferentially recycled back to the cell surface while when an EGF/EGF-R complex were internalized in the same cell type, both components were efficiently degraded (Ebner and Derynck (1991), Cell Regul.2, 599-612). These results suggested that the differences in biological activity of each growth factor might be due to differences

in the intracellular trafficking mechanisms. On the other hand TGF α is a more potent angiogenic factor than EGF (Schreiber et al. (1986), Science 232, 1250-1253.)

Concerning the expression in tumors, there are evidences of the presence of EGF precursor in membranes of some epithelial tumors but TGF α is most express in epithelial tumors , and its action, contrary to EGF, is by an autocrine loop with the EGF-R. On the other hand, results of our center indicate that in some epithelial tumor biopsies there are expression of TGF α and non EGF (breast ductal carcinoma, larynx carcinoma), while other tumors present more EGF than TGF α (non small cell lung cancer (NSCLC)). These results suggest that growth factors can have different impacts in the tumor biology of different neoplastic cells.

All the evidences accumulated in these years about the relationship among the system EGF-R / EGF-R ligands and cancer, convert this system in a very attractive target for the cancer immunotherapy.

Previous results of our group have demonstrated the possibility to develop an active cancer immunotherapy with a vaccine based on EGF. In fact, preclinical and clinical evidences have been obtained about the immunogenicity and low toxicity caused by the vaccination with hEGF coupled to a carrier protein (González et al. (1996), Vaccine Research 5(4), 233-243.)

Preclinical studies have shown that immunization of mice with hEGF in adjuvant increases the survival of mice transplanted with Ehrlich ascites tumor (EAT) (González et al. (1996), Vaccine Research 5(4), 233-243.).

A fusion protein between hEGF and P64k was produced. This protein contains the hEGF sequence inserted between amino acids 45/46 of P64k. This fusion protein was used to immunize mice, causing a specific humoral immune response against hEGF. The immune response generated provoke an increase of life span of EAT bearing mice (González and cols (1997), Vaccine Research 6(2), 91-100)

In two pilot clinical trials with patient of NSCLC, was observed an trend to increase the survival in vaccinated patients compared with an historical control. In patients with high antibody response against the hEGF a marked increase of survival was observed (González et al. (1998), Annals of oncology 9, 1-5.).

In general the vaccination with EGF doesn't generate a specific antibody response against TGF α . However have been obtained evidences that the vaccination with an immunogenic preparation containing TGF α in a murine model generates low levels of anti-EGF antibodies only in some mice. This antibody response in some cases is able to block the EGF binding to its receptor *in vitro*. However the levels of anti-EGF antibodies obtained are not enough to generate an effective EGF immune-castration response with impact in the anti-tumoral action.

Because the action of each of these growth factors is different in each tumor and/or between the primary tumor and its metastasis, a vaccine that combines the two main ligands of the EGF-R, TGF α and EGF, have a better anti-tumoral effect in epithelial tumors, in general sense.

Until the moment of the present invention, has not been developed any therapy that proposes the use of a vaccine preparation containing hTGF α or any derived, or a combination with other ligand of the EGF-R, EGF, in the active cancer immunotherapy.

The present invention claims a vaccine composition that contains hTGF α or any derived of any source, bound genetically (fusion protein) or coupled by chemical methods to a carrier protein, able to inhibit the growth of epithelial tumors without adverse collateral effects. This action is through a growth factor immune-castration mechanism. It also claims a vaccine composition that contains combination of hTGF α with hEGF or any derived together with a carrier protein.

The vaccine composition will be able to be used in the treatment of epithelial tumors dependent of TGF α or TGF α /EGF, or in any other disease associated with TGF α such as psoriasis (Kapp et al (1993) J Dermatol Sci, Jun;5(3):133-42).

In the specification of TGF α , any fragment derived of TGF α that has the same immunology properties and/or similar effects to the original molecule is included. Those derived include, but they are not excluded other, original substitutions of amino acids, change of specific amino acids that increase the stability and/or the activity, chemical modifications, among others.

VACCINE COMPOSITION CONTAINING TRANSFORMING GROWTH FACTOR ALPHA (TGF α). IT USE IN MALIGNANT DISEASES THERAPY.

5 DETAILED DESCRIPTION OF THE INVENTION

The invention consists on a vaccine composition able to cause an immune-castration of self-TGF α that can be used for the treatment of certain cancers and other diseases related with TGF α .

10 On the other hand this invention include the use of a vaccine preparation constituted by a combination of TGF α and EGF. This vaccine can be used for the treatment of neoplasias that depend on these two growth factors in the course of its pathogenesis.

1 - Immunogenic preparations:

15 In the present invention a vaccine preparation used includes the hTGF α either coupled with a carrier protein for methods of genetic engineering (fusion protein) or for chemical methods of conjugation. The hTGF α used in anyone of these immunogenic preparations can be recombinant, synthetic or obtained from natural source. Different proteins can be used as carriers. As examples of carrier proteins can be used: Toxoide Tetanic, KLH, and P64k protein from *Neisseria meningitidis*, among others. The optimum quantity of hTGF α in the vaccine formulation oscillates between 5 μ g and 1000 μ g per dose.

20 On the other hand a vaccine preparation that contains a combination of hTGF α with hEGF (Office of National Registration of Medications, HEBERMIN Not 1266) is used.

25 In the specification of TGF α or EGF, any fragment derived from TGF α or EGF that has the same immunology properties and/or similar effects to the original molecule is included. Those derived include, but they are not excluded other, original substitutions of amino acids, change of specific amino acids that increase the stability and/or the activity, chemical modifications, among others.

A) Obtaining of a fusion protein TGF α -carrier protein by genetic engineering methods:

30 The gene coding for hTGF α (500 pb) was amplified by polymerase chain reaction (PCR) using specific primers. The resulting DNA fragment is digested and cloned in a specific site to an expression vector containing the gene coding for the carrier protein. The resulting protein includes a single or multiple copies of both molecules. You can use an expression vector of mammalian cells, bacteria or yeast. The vector can also include six histidines in the N-terminal end of the carrier protein. The resulting plasmid is verify by restriction analysis on agarosa gels, DNA sequencing using Sequenase 2.0 (Amersham-USB), and finally, analysis of expression of fusion protein in any E.Coli expression strain by Western Blott technique, using an antibody specific monoclonal against hTGF α (R&D System). To obtain the protein the bacterial walls is disrupted using a strong rupture method and then the protein are becomes purified for a combination of differential precipitation methods with ammonium sulfate and chromatography methods. Finally, the protein is filtered under sterile conditions and conserved to - 20°C or lyophilized and conserved at 4°C until its later use.

40 B) Obtaining of a chemical conjugated containing hTGF α :

45 Different preparations that contain hTGF α conjugated with different carrier proteins (as P64k) are obtained. Any chemical conjugation method can be used. As preferential chemical method is used the method using EMCS agent described in the North American patent, U.S.Pat, Not. 4,302,386; Lee et al., 1981.

50 Alternatively, you can use the conjugation method with glutaraldehyde. Briefly, these two or three molecules to a concentration of 1 mg/mL in the final solution are mixing with glutaraldehyde to 0.05% (in the total solution). The mixture is incubated for 1 hour at room temperature and then dialyzed against a solution of PBS 1X/10 mM MgCl₂. Finally, a dialysis against PBS 1X is carried out overnight at 4°C. The immunogenic preparation is filter under sterile conditions and stored at 4°C until its use.

C) Obtaining of a vaccine that combined hTGF α and hEGF.

The obtaining of a vaccine that combines the two main ligands of the EGF-R can be performed in different ways:

1 - Mixing the two vaccines that contain hTGF α or hEGF for separate linked to a carrier protein in a relationship 1:1 just in the moment of the injection. For this purpose can be used the fusion proteins or those chemistry conjugated of each growth factor and carrier protein. The optimum quantity of hTGF α and hEGF in the vaccine formulation oscillates between 5 μ g and 1000 μ g per dose.

2 - Obtaining of a similar genetic construction to the one described in the section A containing both growth factors, hTGF α and hEGF or a combination of anyone of their derived.

3 - Chemical obtaining of a chemist conjugated containig hTGF α and hEGF or a combination of anyone of their derived and a carrier protein using the methodology described in the section B.

D) Obtaining of the immunogenic preparation:

To obtain the desired immunogenic effect of the vaccine compositions is convenient to use an appropriate adjuvant and to select an administration route in which the vaccine preparation exhibits a high immunogenicity.

The vaccine compositions referred in this invention are prepared in two specific ways:

1) Using Al(OH)₃ as adjuvant to obtain a watery solutions from the vaccine preparation adsorbed to this compound. For this purpose is used a range from 5 μ g to 1000 μ g of TGF α equivalent in the different preparations bound to a range from 2 to 5 mg of Al(OH)₃. This formulation is left in agitation for 1 hour. The final solution is conserved at 4°C until its later use.

2) Using incomplete adjuvant of Freund that allow the formation of aqueous/oil or oil/aqueous emulsions. The quantities of fusion protein or chemist conjugated and adjuvant in the final formulation are in a range from 40/60 to 60/40 (volumen/volumen). The volumes depend on the final emulsion desired. The adjuvant is added before the immunization and the formulation is agitated for a period from 10 to 30 minutes, at room temperature.

The final volumes of each immunogenic preparation cover the appropriate range for the corresponding route of administration.

In the case of the combined vaccines prepared just in the moment of the injection as was described in the section C , the two vaccine mixed with the appropriated adjuvant like it has been described previously can be mixed by agitation and inject or inject separate.

The administration of the vaccine compositions can be done for diverse routes: intramuscular, subcutaneous, intranasal and intracutaneous.

EXAMPLES

Example1: Obtaining the DNA segment coding for mature TGF α by polymerase chain reaction (PCR).

The gene coding for hTGF α was amplified by PCR using as template the PSK/TGF α vector (CIGB, Cuba). That plasmid contains the hTGF α complementary DNA (cDNA) cloned in the EcoR V site of commercial vector pBluescript KS - (Stragene). The sequence coding for the mature TGF α (50 amino acids long (Fig.1)) was amplified using the specific primers describe above:

N-Terrminal: 5' - GCTCTAGAAAGTGGTGTCCCATTTTAATGAC-3'

(Underlined, XbaI restriction site)

C-terminal: 5' -CGGAATTCGCCAGGAGGTCCGCATGCTCAC-3'

(Underlined, EcoRI restriction site)

Briefly, 200 ng of the PSKTGF α was used in 75 μ L of a mixture that contains: 500ng of each one of the specific primers, a mixture of deoxynucleotide triphosphates to a concentration of 200mM each one, 25 mM MgCl₂ and 100 Units of TaqPolimerasa enzyme (Promega) in buffer solution given by Promega. A protocol of 30 cycles of denaturalization (1 minute at 94°C), annealing (1 minute at 60°C), and extension (30 seconds at 72°C) was followed. Before the first cycle, DNA was denatured for 4 minutes and after the last cycle; a final extension of 2 minutes was performed.

PCR product is electrophoretically separate on low gelling temperature (LGT) agarosa gels and the amplified gene segment becomes purified according to conventional procedures of extraction with phenol and enzymatic digested using Xba I and EcoR I enzymes (NEB, USES). Following this protocol is prepared the gene segment coding for the mature TGF α .

Example 2: Obtaining of the expression vector for the fused protein TGF α -P64K.

The expression vector pM 92 was used (CIGB, Cuba). It plasmid contains the *lpdA* gene coding for P64k protein from *Neisseria meningitidis* (strain B385) under the control of E.Coli tryptophan operon promoter (*ptrp*) and phage T4 transcriptional terminator (*tT4*). pM 92 code for ampicillin and kanamycin antibiotic resistance. A Dam - E.Coli-strain (GC-366) is transformed with pM92 and the plasmid is purified using a commercial kit (Quiagen) according to the protocol of the manufacturer. PM92 vector were digested and purified from LGT agarosa gels. Subsequently PM92 vector is ligated with the cDNA from mature hTGF α previously prepared, using T4 ligase enzyme (Gibco BRL). The resulting plasmid pMTGF α codes for the fusion protein that contains hTGF α inserted among the amino acid 45/46 of P64k. This recombinant plasmid was verified by restriction analysis in agarosa gels, DNA sequencing using Sequenase 2.0 (Amersham-USB), and finally, analysis of the fusion protein expression in E.Coli MM299 strain by Western Blotting technique using a monoclonal antibody specific for hTGF α (R&D System). The figure 2 shows a schematic representation of the expression vector pMTGF α obtaining process. This vector codes for the fusion protein TGF α -P64K.

Example 3: Obtaining of the expression vector of fused protein TGF α -P64K with six histidines in the N-terminal end (pMHisTGF α).

The expression vector pMHisTGF α was obtained following the same protocol described in the previous example using as starting vector pM224, that includes a segment coding for six histidines in the N-terminal end of P64k. The six histidines tag present advantages in the purification of the protein because increment the binding to chelating Sepharose charged with Cu²⁺ or other metals.

Example 4: Fusion protein (TGF α -P64K) purification.

E.Coli bacteria (strain MM299) expressing the fusion protein TGF α -P64K were grown in LBA medium (10 g/L Triptona, 5g/L Yeast Extract, 10 g/L NaCl and 50 mg/L ampicillin) for 10 hours at 37°C. After cells collection, all steps were performed at 0-4°C. Bacterial disruption was achieved in a French press at 1500 kg/cm², and the insoluble fraction was removed by high-speed centrifugation for 30 minutes at 11,000xg. As first purification step a 40% of ammonium sulfate precipitation was done to remove part of the E.Coli proteins. The resulting pellet was removed by a further centrifugation at 4°C for 30 minutes to 11,000 x g. The supernatant was fractioned by hydrophobic interaction chromatography (TSK-butyl, Pharmacia, Sweeden), with a decreasing gradient of ammonium sulfate from 40% to 0% in buffer Tris-Cl, pH = 7.2 containig 0.15M NaCl. Subsequently the resulting sample was separated by gel filtration on a G200 column (Pharmacia) equilibrated with PBS 1X, achieving a final purity of more than 95%. Proteins concentration is determined using a colorimetric method described by Lowry et al (1951) J.Biol.Chem. 191, 495-498). The characterization of fused protein was done by Western Blotting technique, using specific antibodies against P64k and TGF α .

Example 5: Fusion protein (HisTGF α -P64k) purification.

E.Coli bacteria (strain MM299) expressing the fusion protein TGF α -P64K were grown in LBA medium (Triptona 10 g/L, Yeast Extract 5g/L, NaCl 10 g/L and 50 mg/L ampicillin) for 10 hours at 37°C. After cells collection, all steps were performed at 0-4°C. Bacterial disruption was achieved in a French press at 1500 kg/cm², and the insoluble fraction was removed by high-speed centrifugation for 30 minutes at 11,000xg. As first purification step a 40% of ammonium sulfate precipitation was done to remove part of the E.Coli proteins. The resulting pellet was removed by a further centrifugation at 4°C for 30 minutes to 11,000 x g. The supernatant was fractioned by chelating affinity chromatography (Chelating Sepharose Fast Flow, Pharmacia, Sweeden), due to the presence of six histidines in the protein, with a increasing Imidazol gradient from 25 mM to 500 mM in buffer Tris-Cl, pH = 5.5 containig 0.5 M NaCl. Subsequently the resulting sample was passed through on gel filtration G25 column (Pharmacia) equilibrated with PBS 1X to remove the salts, achieving a purity level of 95%. Proteins concentration is determined using a colorimetric method described by Lowry et al (1951) J.Biol.Chem. 191, 495-498). The characterization of the fused protein was done by Western Blotting technique, using specific antibodies against P64k and TGF α .

Example 6: Recognition of the recombinant protein TGF α -P64K for a monoclonal antibody (Mab) specific for hTGF α .

To determine if TGF α could be recognized by an anti-hTGF α Mab (Calbiochem) in the fused protein context, was done a Western Blotting technique. Electrophoretically 25 μ g of EGF-P64K, TGF α -P64K or P64K were separated in two polyacrilamide gels and then transferred to a 0.45 μ m nitrocellulose membrane according to conventional procedures. After the transfer, membranes were incubated with a blocking solution of TBS 1X with 5% of skim milk overnight at 4 °C. After a brief wash with TBS 1X-Tween 20 (0.05%), membranes were incubated, one replies with an antibody anti-P64K (1 / 500) (Fig.3A) and the other one with a anti-TGF α Mab (1/100) (Fig.3B) for 2 hours at room temperature. Subsequently were performed 3 washes with the same solution and membranes were incubated with alkaline phosphatase-labeled goat anti-mouse immunoglobulins (1/1000) for 1 hour in same conditions. Finally was added 0.004 g of Fast Net enzyme substrate (Sigma) in buffer containig 0.1 M Tris-Cl pH=8.2, 0.004 g of Naphtol ACE-MX Phosphate (Sigma) and 400 μ L of NN'Dimetil Formamide in 20 mL. The reaction stopped with similar washes. A specific recognition of TGF α -P64k by the anti-hTGF α Mab was observed (Fig. 3). This result demonstrates that TGF α in the fusion protein maintains a structure able of being recognized by a specific antibody.

Example 7: Obtaining of a chemical conjugated hTGF α -P64k.

A milliliter of TGF α in PBS/10mM MgCl₂ at 2 mg/mL is mixed with a milliliter of P64k at 2 mg/mL in the same solvent. Then 0.2 mL of 0.5% glutaraldehyde solution was added for a final percent of 0.05%. The mixture was incubated 1 hour at room temperature, and dialyzed against a PBS 1X/10 mM MgCl₂ solution. Finally, dialysis against PBS 1X was carried out overnight at 4°C. The immunogenic preparation is filtered under sterile conditions and stored at 4°C until its use.

Example 8: Obtaining of a fusion protein between hTGF α , hEGF and P64k.

The gene coding for hEGF (150 pb) it is amplified by PCR using the plasmid pBEF 10 as template. That plasmid contains the complete hEGF cloned in the EcoR V site of commercial vector pBluescript SK II (Stragene). The obtained DNA is linked to the pMHisTGF α plasmid in a Bam HI site located in the C-terminal end of the P64k using the methodology described in the example 2. This way the pMTGF α -EGF vector is obtained that codes for the fusion protein TE-P64k.

Example 9: Obtaining of a chemical conjugated hTGF α -hEGF-P64k.

A milliliter of TGF α in PBS/10 mM MgCl₂ at 3 mg/mL is mixed with a milliliter of hEGF at 3 mg/mL and P64k at 3 mg/mL in the same solvent. Then 0.6 mL of 0.5% glutaraldehyde solution was added for a final percent of 0.05%. The mixture was incubated 1 hour at room temperature, and dialyzed against a PBS 1X/10mM MgCl₂ solution. Finally, dialysis against PBS 1X was carried out overnight at 4°C. The immunogenic preparation is filtered under sterile conditions and stored at 4°C until its use.

Example 10: Preparation of formulations that contain hTGF α .

The different immunogenic preparation described in the examples 2, 3, 7, 8 and 9 are mixed with Al(OH)₃ or Montanide ISA 51 as was described in detailed description of the invention. Quantities used that are equal to 50 μ g of hTGF α in all the preparations and 50 μ g hTGF α and hEGF, in the case of the combined vaccines described in the examples 8 and 9. Two milligrams of Al(OH)₃ was used by each preparation of fused protein or chemist conjugated containing respectively 50 μ g hTGF α or hEGF equivalent.

Example 11: Preparation of a combined vaccine containing TGF α -P64K protein and EGF-P64K protein.

Fifty micrograms of each growth factor in 0.6 mg of recombinant are mixed in a total volume of 0.5 mL with same volume of Montanide ISA 51 and agitated for 10 minutes at room temperature before the injection.

In the case of using Al(OH)₃ as adjuvant, two preparations containing 0.6 mg of each protein in 2 mg of Al(OH)₃ are mixing before the injection.

Example 12: Preparation of a combined vaccine containing a chemical conjugated TGF α /P64K and EGF/P64K.

An immunogenic preparation containing 50 µg of hTGFα linked to P64k as described in the example 7 in 0.25 mL is mixed with 0.25 mL of an equals immunogenic preparation that contains hEGF and mixed with 0.5 mL of Montanide ISA 51 according was described in the example 10, using a syringe, for a period of 10 minutes at room temperature.

- 5 In the case of using Al(OH)₃ as adjuvant, 0.5 mL of each one of those chemical conjugated described before that contain 50µg of hTGFα or hEGF respectively adsorbed in 2 mg of Al(OH)₃ are mixed.

Example 13: Immunogenicity of TGFα-P64K / Incomplete Freund adjuvant (Montanide ISA 51) in a murine model.

- 10 To demonstrate the immunogenicity of the vaccine, Balb/c mice, females among 6-8 weeks, were injected subcutaneous with 58 mg (5µg equivalent of TGFα), 116mg (10µg) or 0,6mg (50µg) of TGFα-P64k with Montanide ISA 51 in a 1:1 proportion. The immunogen was prepared as described in the detailed description of the invention and agitated for 10 minutes before the immunization. Each animal received 4 doses. The blood was extracted before the immunization, one week later and
- 15 biweekly since that moment. The serum was separated from the extracted blood of animals and specific antibodies titer was determined against the hTGFα by an indirect ELISA technique.

- Briefly, microtiter ELISA's plates (COSTAR) were coated with 50µL/well of an hTGFα solution at 2.5 µg/mL in buffer carbonate-bicarbonate pH= 7.2 and incubated overnight at 4°C. After three washes with PBS 1X-Tween 20 (0.05%), the plates were blocked with a solution of PBS 1X-Tween 20 (0.05%)
- 20 -SFT (5%) for 1 hour at 37°C. Immediately the serums of the immunized mice were added and incubated for 2 hours at 37°C. After washing, the plates were incubated with alkaline phosphatase-labeled goat anti-mouse immunoglobulins (Sigma) diluted 1/1000 in PBS 1X-Tween 20 (0.05%) -SFT (5%) (50µL /pozo) for 1 hour at same temperature. Finally, after washing, the substrate of the enzyme (p-nitrophenilphosphate(Sigma)) was added to final concentration of 1 mg/mL in buffer Dietanolamine pH=9.8 (50µL /well). The absorbance at 405 nm of enzyme-substrate complex formed was measured in an ELISA plate reader.

- 25 The figure 4 show the kinetics of the polyvalent anti- hTGFα antibody response obtained in mice immunized with TGFα-P64K.

- 30 Due to the high homology between hTGFα and rat or mouse counterpart (93%) you can consider this immune response against hTGFα as a response against self-TGFα (murine).

Example 14: Immunogenicity of TGFα-P64K / Al(OH)₃ in a murine model.

- 35 An immunization protocol was done according was described in the previous example, using 2 mg Al(OH)₃ as adjuvant. The immunogenic preparation got ready according was described in the detailed description of the invention. Antibody titers up to 1/10000 were obtained for TGFα. The technique used to determine the antibody titers was the indirect ELISA described in the example 13.

Example 15: IgG Subclass distribution in mice immunized with TGFα-P64k protein / incomplete Adjuvante of Freund (Montanide ISA 51) in a murine model.

- 40 The IgG subclass distribution was determined by an indirect ELISA'S technique described in the example 13, using specific antiserum against the different IgG subclasses conjugated with biotin (Jackson) to a dilution of 1/1000 and later on the complex streptavidine-phosphatase (1/1000). The proportion of each IgG subclass was determined regard to total IgG in the serum of immunized animals. The animals were immunized with 50 µg of TGFα in the fused protein using subcutaneous
- 45 (Group 1) or intramuscular (Group 2) route following the immunization protocol described in the example 13. In the figure 5 is observed the subclass distribution obtained. A bigger proportion of IgG1 was obtained in both groups of mice used in the study.

- 50 Example 16: Determination of the capacity of animal serums immunized with TGFα-P64k protein / incomplete Adjuvante of Freund (Montanide ISA 51) of inhibit the binding of I₁₂₅-TGFα by radio receptor assay technique (RRA).

To determine if the antibodies generated in the immunization protocols described previously were able to inhibit the TGFα binding to its receptor, was done an in vitro technique called RRA. In synthesis, the serums of immunized mice obtained as it is described in the example 13 were incubated with a

mixture that includes 100 mL of human placenta membrane and 20 mL of I_{125} -TGF α (100000 cpm) and 330 mL of buffer: 10 mM Tris-Cl, 10 mM MgCl₂ and BSA to 1%, pH=7.4. TGF α was coupled to ^{125}I radioisotope using the method of chloramine T (Hunter and Greenwood (1962, Nature, 358:495-498). The mixture was incubated by 1 hour at room temperature and the reaction was stopped with 1 mL of the buffer mentioned before. Finally the tubes were centrifuged at 1000 rpm for 30 minutes. Pellets were washed and allowed to dry off. The radioactivity was measured in a gamma emission counter (Wallac, Finland). The decrease in the measured values of radioactivity indicates the inhibition of binding between TGF α and its receptor, due to the action of tested serums. A range of 50%-80% inhibition percent was obtained among all tested serums.

Example 17: Determination of the anti-human EGF (hEGF) humoral response generate by the immunization with TGF α -P64k protein.

The presence of anti-EGF antibodies was determined in serums of mice that showed high anti-TGF α antibodies titers using the described indirect ELISA'S technique. Dilutions of 1/100, 1/1000 and 1/10000 of serums were added to plates coated with hEGF (CIGB). The figure 6 shows the anti-EGF antibody titers obtained in the serum of mice immunized with TGF α -P64k protein. Only in a group of immunized mice a positive anti-EGF antibodies response is obtained.

However mice immunized with one chemical conjugated EGF-P64K didn't show any level of anti-TGF α antibodies.

Example 18: Recognition of human tumors in vitro for a policlonal antiserum anti-hTGF α obtained by the immunization with the TGF α -P64k protein / incomplete Adjuvante of Freund (Montanide ISA 51).

Policlonal antisera anti-hTGF α obtained immunizing mice with TGF α -P64k protein in Montanide ISA 51, was used to determine the TGF α expression in biopsies of tumors included in paraffin. These biopsies were obtained from patients vaccinated with a vaccine based on EGF. In a patient with high anti-hEGF antibodies response, a regression of NSCLC tumor was observed. However, later was detected a second larynx tumor. Biopsies of these two tumors were analyzed and it was observed a differential expression of EGF and TGF α in each one of them. In the figure 7 the reactivity values obtained with the different antibodies are shown. These results confirm the fact that immunization with the vaccine preparation containing TGF α -P64k provokes specific antibodies against hTGF α able to recognize this molecule in human tumors.

Example 19: mRNA expression of EGF, TGF α and EGF-R in breast carcinoma biopsies.

Messenger ribonucleic acid (mRNA) was isolated from breast carcinoma tumor biopsies using TRIZOL reagent (Life technologies) and converted to cDNA by the reverse transcriptase enzyme. The total cDNA underwent 30 cycles of PCR using specific primers for each one of these molecules. As internal control was used a housekeeping gene (GAPDH). The PCR products obtained were separated electrophoretically on 1.5% agarosa gels and visualized with Etidium bromide.

In the figure 8 are shown the results obtained using the specific primers for EGF, TGF α , EGF-R and GAPDH (internal Control) in 22 breast carcinomas. EGF mRNA was observed only in 1/22 biopsies, however it was observed a high expression of TGF α and EGF-R mRNA in most of the samples. The high correlation between the expression of these two molecules, suggests the importance of the autocrine loop TGF α / EGF-R in the growth of this type of tumors (Figure 9).

BRIEF DESCRIPTION OF THE DRAWINGS.

Figure 1: Genetic and amino acidic sequence (letters underlined in boldface) of mature hTGF α .

Figure 2: Schematic representation of expression vector pMTGF α -obtaining process.

Figure 3: Recognition of TGF α -P64K fusion protein by anti-P64K Mab (A) and anti-hTGF α Mab (B) by Western Blotting technique. 10% SDS-PAGE was carried out to P64K (1), EGF-P64K (2) and TGF α -P64K (3). Then proteins were transferred to a nitrocellulose membrane and incubated with specific

antibodies against P64K (A) or TGF α (B) with the objective of characterizing the fused protein between TGF α and P64K.

Figure 4: Anti-hTGF α antibody response kinetics: The specific antibodies titers against hTGF α were measured by indirect ELISA'S technique. The mice were immunized with 5 μ g (A), 10 μ g (B) and 50 μ g (C) of hTGF α -equivalent in TGF α -P64K protein mixed with Montanide ISA 51. The x-axis represent the days when the samples were collect in each mouse and the y-axis the reciprocal of the antibody titter reached. The days of immunization they are pointed out with arrows in the graph A (Day 0, 14, 28 and 42).

Figures 5: IgG subclass distribution induced by the immunization with 50 μ g of TGF α -equivalent in the fused protein. Comparison of IgG subclass proportion in the antibody response induced with the immunization of TGF α -P64k protein subcutaneous (1) or intramuscular (2). The values of standard deviation are shown in the figure for each one of the groups of 5 immunized animals.

Figures 6: Anti-EGF specific antibodies response in mice immunized with the TGF α -P64K fused protein. In the chart are shown anti- hTGF α and anti-EGF antibody titers reached in mice immunized with TGF α -P64K.

Figures 7: Determination of EGF-R, EGF and TGF α expression by immunohistochemistry in tumors biopsies of vaccinated patients included in the pilot II clinical trial of EGF-vaccine. The differential reactivity of these three molecules in the primary tumor of lung and of a second primary tumor of larynx that appeared after, are shown with positive signs in the figure.

Figures 8: EGF, hTGF α and EGF-R ARNm expression in 22 breast carcinomas. The figure show the products of 30 cycles PCR obtained with specific primers for each molecule and visualized with etidium bromide after being separated on 1.5% agarosa gels. GAPDH ARNm expression, used as internal control, is also observed.

Figures 9: Correlation among the hTGF α and EGF-R ARNm levels in breast carcinoma biopsies: The bands intensity obtained with etidium bromide was analyzed by means of a calculation program (ImagQuant, Amersham). The x-axis shows relation between the intensity values for the PCR products using specific primers for EGF-R and those obtained with GAPDH for each sample (relative intensity) and the y-axis the same relative intensity value for hTGF α . A positive correlation was observed between the expressions of these two molecules ($R^2 = 0.657$, $p=0.00121$).